Supplementary Information

Combined physical and biological contributions to radiotherapy enhancement by Lu-based nanoscintillators in pancreatic cancer models

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Intrinsic Radiosensitivity of PANC-1 vs MIA PaCa-2 cell lines



Figure S1: Survival fraction (mean \pm SEM) of PANC-1 (pink) and MIA PaCa-2 (blue) cells plotted as a function of irradiation dose, normalized to the 0 Gy condition. Data was fitted to a linear quadratic model using Prism and significance was calculated using a two-way ANOVA. Results were collected from 6 wells per condition from at least 2 independent experiments. (*) indicates p < 0.05, (**) indicates p < 0.01, (***) indicates p < 0.001, and (****) indicates p < 0.001.

γ-H2AX foci in PANC-1 cells after X-ray irradiation



Figure S2: Representative microscopy images showing the **A**) γ -H2AX signal, **B**) Hoechst signal, and **C**) merged images of PANC-1 cells collected 1 hour after X-ray irradiation. In images i-iii, immunofluorescence staining for γ -H2AX foci was performed on PANC-1 cells after they received 0, 2, or 2.8 Gy of X-rays. In images iv-v, cells were first incubated with 0.5 mg/mL Lu₃Al₅O₁₂@SiO₂ for 24 hours, then received either 0 or 2 Gy of X-rays. Scale bar = 25 μ m.



Figure S3: Representative microscopy images showing the A) γ -H2AX signal, B) Hoechst signal, and C) merged images of PANC-1 cells collected 24 hours after X-ray irradiation. In images i-iii, immunofluorescence staining for γ -H2AX foci was performed on PANC-1 cells after they received 0, 2, or 2.8 Gy of X-rays. In images iv-v, cells were first incubated with 0.5 mg/mL Lu₃Al₅O₁₂@SiO₂ for 24 hours, then received either 0 or 2 Gy of X-rays. Scale bar = 25 μ m



γ-H2AX foci in MIA PaCa-2 cells after X-ray irradiation

Figure S4: **A**) and **C**) show representative microscopy images taken of MIA PaCa-2 cells collected either 1 hour or 24 hours post-irradiation, respectively. In images i-iii, immunofluorescence staining for γ -H2AX foci was performed on PANC-1 cells after they received 0, 2, or 2.8 Gy of X-rays. In images iv-v, cells were first incubated with 0.5 mg/mL Lu₃Al₅O₁₂:Pr³⁺@SiO₂ for 24 hours, then received either 0 or 2 Gy of X-rays. Scale bar = 25 µm. The number of foci per nucleus was quantified, and the frequency distributions for each condition are shown in panels **B**) and **D**) for cells collected 1 hour and 24 hours after irradiation, respectively. The average number of foci per nucleus (mean ± SEM) in cells irradiated with 2 Gy of X-rays is shown in graphs **E**) and **F**) for samples collected 1 or 24 hours post-irradiation, respectively. **G**) presents the fraction of foci remaining at 24 hours post-irradiation normalized to the number of foci present at 1 hour post-irradiation for the 2 Gy, 2.8 Gy, and 2 Gy + Lu₃Al₅O₁₂:Pr³⁺@SiO₂ conditions. Foci quantification

data was taken from at least 80 nuclei per condition. Statistical significance was assessed using a two-way ANOVA followed by Tukey post hoc test. (*) indicates p < 0.05, (**) indicates p < 0.01, (***) indicates p < 0.001, and (****) indicates p < 0.0001.

Efficacy in 3D models of pancreatic cancer under monochromatic synchrotron irradiation



MIA PaCa-2



Figure S5: Viability (A-D) and area (E-H) of PANC-1 (A-B, E-F) and MIA PaCa-2 (C-D, G-H) spheroids after irradiation with monochromatic synchrotron radiation of 62.31 keV (B, F, D, H) or 64.31 keV (A, E, C, G). Data was collected 6 days post-irradiation and is presented as mean \pm SEM. The results were normalized to the 0 Gy condition and fitted with a nonlinear regression in Prism according to the [inhibitor] versus response model with three parameters. Statistical significance was assessed using a two-way ANOVA followed by Tukey post hoc test. (*) indicates p < 0.05, (**) indicates p < 0.01, (***) indicates p < 0.001, and (****) indicates p < 0.0001.