SUPPLEMENTARY FIGURES

SUPPLEMENTARY FIGURE 1

A) Nichoids’ quality control images obtained by Scanning Electron Microscopy (SEM) at 5kV. SEM images of: a single niche (Scale bar: 30μm), a block of 5x5 of single niches (Scale bar: 100μm), and a supermatrix of niches (Scale bar: 300μm). The images show a representative example of a good manufacture result used for biological experimental validation.
B) Schematic representation of the new optimized set-up for the multi foci laser polymerization composed by femtosecond laser source, a beam expander, a 4f telescope and a Spatial Light Modulator (SLM).
SUPPLEMENTARY FIGURE 2

a

NESTIN/DAPI  GFAP/DAPI  MAP2/DAPI  DCX/DAPI

b

NPCs  GFP NPCs

Total number of spheres/well (diameter 70-100 μm)

NPCs  GFP NPCs

7 days in culture

Proliferation curve

Number of cells

10^5/well at plating time

3  5  7 days

NPCs  GFP NPCs

7 days in culture

c

d

GFAP/Nestin/DAPI  GFAP/Map2/DAPI  GFP/Nestin/DAPI  GFP/Map2/DAPI  GFP/GFAP/DAPI  GFP/GFAP2/DAPI

Number of surviving cells

Nestin  Map2  GFAP  NO2
SUPPLEMENTARY FIGURE 2. Characterization of NPCs obtained from the SVZ

A) Representative direct live image of the Sub Ventricular Zone (SVZ) from where NPCs were isolated. Scale bar 100 μm. The adjacent panels show representative SVZ-immunofluorescence characterization evaluating the expression of NESTIN (red), MAP2 (red), GFAP (red), DCX (red) and DAPI for nuclei (blue). Scale bar 20 μm.

B) Representative direct live images of NPCs and GFP NPCs after 7 days of culture in floating conditions. Scale bar 200 μm. The histograms report the evaluation of the number of spheres with a dimension comprised between 70-100 μm diameter and the proliferation curve respectively of NPCs and GFP NPCs seeded at a starting density of 1x10⁴ cells. Cells were counted in triplicate with trypan blue exclusion method after 3, 5, and 7 days. Data are reported as mean ± SD.

C) Representative immunofluorescence images of neurospheres [NPCs (top panels) and GFP NPCs (lower panels)] expressing NESTIN (red) and MAP2 (red). The lower panels also show co-staining of GFP positive cells (green). DAPI (blue) was used for nuclei staining. Scale bar 100μm.

D) Representative immunofluorescence images of GFP NPCs differentiated for 7 days (See Materials and Methods section for further details). Cells were characterized by evaluating the expression of NESTIN (red), MAP2 (red), GFAP (red) and NG2 (red). Nuclei were labeled in blue (DAPI). The histogram reports the percentage of GFP NPCs positive to the respective markers. Data are expressed as mean of the quantification performed in three different fields for each condition ± SD (n=3). Scale bar 20 μm.
SUPPLEMENTARY FIGURE 3

A) In vivo direct light images (EVOS FL microscope, Euroclone) of NPCs neurospheres maintained in stem cells medium in standard floating conditions (CONTROL) or grown inside the Nichoid (NICHOID) at day 3, 7, 10 and 14. The images refers to NPCs and GFP NPCs.

SUPPLEMENTARY FIGURE 3. Proliferation of NPCs expanded inside the Nichoid
Scale bar 400 μm. Images are representative of what was observed in at least three different independent experiments.

B) Percentage of dead cells at different time points of NPCs and GFP NPCs grown inside the Nichoid and in standard floating condition. Cells were plated in NSC medium at the density of 1x10^4 cells at plating time. Cells were counted with trypan blue exclusion method after 3, 7, 10, 14 days. Data are expressed as mean of at least three independent experiments ± SD (n=3). Statistical analysis was performed with Student’s t-test followed by Bonferroni post-test (***p<0.001 vs Control NPCs).
SUPPLEMENTARY FIGURE 4

**SOX2** (Mw 34 kDa – Sample 1)  
Abli: anti-rabbit 1:5000

**β-ACTIN** (Mw 42 kDa – Sample 1)  
Abli: anti-rabbit 1:5000

**NANOG** (Mw 35 kDa – Sample 3)  
Abli: anti-rabbit 1:5000

**β-ACTIN** (Mw 42 kDa– Sample 3)  
Abli: anti-rabbit 1:5000

**OCT4** (Mw 45 kDa – Sample 2)  
Abli: anti-rabbit 1:5000

**β-ACTIN** (Mw 42 kDa – Sample 2)  
Abli: anti-rabbit 1:5000

**TUJ1** (Mw 50 kDa – Sample 3)  
Abli: anti-mouse 1:5000  
This sample is the previous one used with anti-SOX2, washed in TBS and then stripped for 10 min at RT

**β-ACTIN** (Mw 42 kDa– Sample 3)  
Abli: anti-rabbit 1:5000

SUPPLEMENTARY FIGURE 4. Full blots of Western Blot analysis.
SUPPLEMENTARY FIGURE 5

SUPPLEMENTARY FIGURE 5. Structural organization and differentiation capabilities of NPCs replated after Nichoid-growth.
A) The box plots report the different dimension of the neurospheres grown inside the Nichoid and in control floating conditions. The histogram reports the percentage of spheres dimension distribution (*p < 0.05 vs Control, **< 0.001 and ***p<0.001 vs Control). Representative direct light images of neurospheres obtained from NPCs grown inside the Nichoid and then replated in standard floating condition (1x10⁴ cells at plating time). Scale bar: 400 µm.

B) Representative direct light images of NPCs expanded inside the Nichoid for 7 days and then differentiated in standard conditions for 7 more days. Cells were plated at the density of 1x10⁴/well. Scale bar: 200 µm.

C) Representative immunofluorescence images of NESTIN, GFAP, BETA-TUBIII, MAP2, and NG2 expression in NPCs grown for 7 days inside the Nichoid and then differentiated for 7 more days. Nuclei are stained in blue (DAPI) and the other markers are in red.

D) Representative immunofluorescence images of MAP2 expression in NPCs grown for 7 days inside the Nichoid and then differentiated for 7 more days in standard conditions. Nuclei are in blue (DAPI) and MAP2 is shown in red. Scale bar 100 µm.
SUPPLEMENTARY FIGURE 6. Experimental set up of in vivo experiments. Experimental set up to investigate in vivo the therapeutic effects of NPCs grown inside the Nichoid.
SUPPLEMENTARY FIGURE 7

Representative immunofluorescence images of ki67 expression in striatal coronal sections of MPTP injected mice treated with Nichoid expanded NPCs. Ki67 is shown in red, Nuclei are in blue (DAPI), green refers to engrafted GFP NPCs Scale bar: 20 μm.
**SUPPLEMENTARY TABLE 1. List of Primers used for Real Time PCR**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward Primer</th>
<th>Reverse Primer</th>
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<tr>
<td>Sox2 FW</td>
<td>CATGTATAACATGATGAGACGGAGCTGAA</td>
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<td>CCTGGAAGATGGTGAGGCTCCCTGTTGA</td>
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**SUPPLEMENTARY TABLE 2. Deregulated genes involved in pluripotency**

Genes were ranked according to their fold change expression in Nichoid versus Control conditions.