

Nanoliposomes for Safe and Efficient Therapeutic mRNA Delivery: A Step Toward Nanotheranostics in Inflammatory and Cardiovascular Diseases as well as Cancer

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Abstract

Rationale: Genetic therapy using modified mRNA for specific therapeutic protein expression for disease treatment and vaccination represents a new field of therapeutic and diagnostic medicine. Non-viral vectors transfection using biocompatible nanoliposomes enables safe and efficient delivery of therapeutic mRNA.

Objective: Generation of non-toxic, cell-compatible cationic nanoliposomes as nanotheranostic agents to successfully deliver therapeutic mRNA.

Methods and results: Cationic nanoliposomes (DC-Cholesterol/DOPE) were generated as transfection vehicles for either eGFP mRNA or the therapeutic anti-inflammatory, CD39 mRNA. We observed no toxicity using these nanoplexes and noted high cell viability after transfection. Nanoplexes for the transfection of eGFP mRNA showed an increase in fluorescence signals on microscopy as compared to the mRNA control after 24 hours in Chinese hamster ovary (CHO) cells (14.29 ± 5.30 vs. 1.49 ± 0.54 ; mean \pm SD respectively; $p < 0.001$) and flow cytometry (57.29 ± 14.59 vs. 1.83 ± 0.34 ; % mean \pm SD; $p < 0.001$). Nanoplexes for the transfection of CD39 mRNA showed increased CD39 expression in flow cytometry (45.64 ± 15.3 vs. 3.94 ± 0.45 ; % mean \pm SD; $p < 0.001$) as compared to the mRNA control after 24 hours using CHO cells. We also demonstrated efficient transfection across several cell lines (CHO, HEK293, and A549), as well as long-term protein expression (120 h and 168 h) using these nanoplexes.

Conclusions: We have developed and tested non-toxic, safe, and efficient nanoliposome preparations for the delivery of therapeutic mRNA that hold promise for novel therapies in diseases such as inflammatory and cardiovascular diseases, as well as cancer. We have also demonstrated that this approach provides a reliable technology to deliver CD39 mRNA as an anti-inflammatory therapeutic for future nanotheranostics approaches.

Key words: anti-inflammatory, mRNA, nanoliposomes, therapeutics, transfection.

Introduction

Over the past years, modified messenger RNAs (mRNAs) have attracted major attention for therapeutic applications in inflammatory and

cardiovascular diseases, in cancer, and for vaccination.¹ The delivery of synthetic mRNA instead of deoxyribonucleic acid (DNA) to targeted cells for

the expression of therapeutic proteins represents a new and exciting field of personalized medicine.²⁻⁴ Therapeutic mRNA offers higher safety through avoiding the need for genomic insertion, and is less immunogenic, and the therapeutic proteins are expressed under physiological conditions by the cells' own translational machinery.^{3,5}

Furthermore, transient expression of the therapeutic proteins allows for a safe approach to be used in patients and is more desirable for some clinical disorders, and as a vaccination vector.^{4,5} Non-viral gene carriers, such as liposomes are cost efficient and highly suitable for non-toxic *in vitro* and *in vivo* delivery of exogenous genetic material and drugs into specific cell types.⁶

Liposomes are small, spherical amphiphilic vesicles consisting of a lipid bilayer, are biodegradable, biocompatible, non-pathogenic, and exhibit high agent-loading efficiency and great diversity in the lipid composition, as well as the possible surface modifications.^{5,7,8} These liposomes are able to entrap hydrophobic agents within the bilayer, which shields the encapsulated agent or drug from degradation through, for example, nucleases.⁹ Cationic liposome-mediated transfection displays the most commonly used and most successfully applied method for the cellular transfection of DNA and siRNA.^{6,10-12} These charged liposomes consist of a cationic lipid and a neutral phospholipid, which can be loaded via electrostatic interactions with nucleic acids due to their negatively charged phosphate groups.^{13,14} These so-called lipoplexes can be taken up by the cells via endocytosis or endocytosis-like mechanisms.^{14,15} The first studies of liposome-based mRNA transfection using cationic DOTMA (1,2-di-O-octadecenyl-3-trimethylammonium propane) / DOPE liposomes have been successful; however, the use of DOTMA is known to show cytotoxicity effects.² Since transfecting mRNA has been attracting major interest for therapeutic purposes, there is a need to develop delivery systems, which possess high transfection efficiency and have no cytotoxic effects.

Here, we present a novel, non-toxic and effective approach for the transfection of mRNA using DC-Cholesterol/DOPE nanoliposomes (NLps). We demonstrate that this innovative approach can be used across several cell lines with good efficiency using the eGFP-encoding mRNA as a proof of concept model. We observed no effects on cell viability or hemocompatibility, highlighting the safety of these NLps. Using this exciting risk-free approach, we also established long term transfection of the anti-inflammatory/anti-thrombotic therapeutic CD39 mRNA.

Material and Methods

Generation of synthetic mRNA

The synthetic mRNAs (eGFP, CD39) were prepared as previously described.^{4,16} The vectors with coding sequences were amplified with a HotStar HiFidelity Polymerase Kit (Qiagen, Germany) using two primers, TTG GAC CCT CGT ACA GAA GCT AAT ACG as the forward primer (Ella Biotech, Germany) and T₁₅₀ CTT CCT ACT CAG GCT TTA TTC AAA GAC CA as the reverse primer (Ella Biotech, Germany). DNA sequences were purified (Qiaquick PCR purification Kit, Qiagen, Germany), and *in vitro* transcription (IVT) was performed using a MEGAscript® T7 Kit (Ambion, Scotland), according to the manufacturer's instructions. The mRNA was modified using the 3'-O-Me-m⁷G(5') ppp(5')G RNA cap structure analog (New England Biolabs, Germany), pseudouridine-5'-triphosphate (TriLink Biotech, U. S. A.), and 5-methylcytidine-5'-triphosphate (TriLink Biotech, U. S. A.). Additionally, an RNase inhibitor (Thermo Scientific, U. S. A.) was added. The IVT was incubated for 4 h at 37 °C.

Afterwards the mRNA was cleaned up with an RNeasy kit (Qiagen, Germany), and eluted in 40 µl nuclease-free water. Next the dephosphorylation of mRNA was implemented with an Antarctic Phosphatase Kit (New England Biolabs, Germany), cleaned up and then eluted in the same amount of nuclease-free water. The concentration of mRNA was confirmed photometrically and the purity of mRNA with 1% agarose gel.

Generation of nanoliposomes and nanoplexes

DOPE (1,2-dioleoyl-sn-glycero-3-phosphoethanolamine) and DC-Cholesterol were purchased from Avanti Polar Lipids (U. S. A.) and used without further purification. The lyophilized lipids were dissolved in chloroform (Sigma-Aldrich, Germany) at 25 mg/ml. Each lipid was further diluted to a convenient working solution of 3 mg/ml. The formulation of 2:1 of DC-Cholesterol and DOPE was used, vortexed, and the solvent evaporated using N₂ gas for several minutes. Residual solvent was removed by vacuum for at least 24 h via the dry film method.^{13,17}

The obtained lipid film was hydrated with 1 ml nuclease-free H₂O by vortexing. After 10 min at room temperature, a cloudy suspension of multilamellar liposomes was sonicated until the solution cleared. The resulting liposomes were reduced in size and sterilized by extrusion about 21 times using an extruder (Avanti, U. S. A.) and a PC membrane 0.1 µm (Avanti, U. S. A.). The resulting cationic nanoliposomes (NLps) were stored in glass vials at 4 °C before

usage. Nanoplexes were formed via incubation of mRNA with NLps at room temperature for 20 min, in either Opti-Mem™ (Gibco, Massachusetts, U. S. A.) or the desired cell media without fetal bovine serum (FBS).

Characterization of nanoliposomes

The size and size distribution of the generated NLps were measured at 25 °C using a Malvern NanoZetaSizer (Malvern Instruments, U. K.) spectrometer (wavelength $\lambda=632.8$ nm). The volume-weighted Gaussian size distribution was fit to the autocorrelation functions, and particle size values were obtained.

Transfection of CHO, HEK293, and A549 cells with generated nanoplexes

For the transfection analysis, HEK293 (human embryonic kidney) cell line, CHO (chinese hamster ovary) cells, and A549 (adenocarcinomic human alveolar basal epithelial) cells were used. HEK293 and CHO cells were maintained in a DMEM (Gibco, U. S. A.), and A549 cells in an RPMI (Gibco, U. S. A.) in a humidified atmosphere of 5% CO₂ at 37 °C. Both media were supplemented with 10% FBS, 1% L-Glutamine, and 1% penicillin and streptomycin.

One day prior to transfection, the respective cells were seeded with a density of 150,000 cells/well onto a 12-well plate (Corning Costar, U. S. A.). The nanoplexes as the transfection solution were prepared fresh as described above. The cells were washed twice with 1 ml PBS (w/o Ca⁺/Mg⁺; PAA, Pasching, Austria), then the transfection was added to the cells and incubated for 24 h, 72 h, 120 h, and 168 h at 37 °C. The incubators used were supplied with 5% CO₂ (Heraeus 6000; Thermo Fischer Scientific, Germany).

XTT-viability assay

Cell viability after transfection was analyzed using XTT (2,3-Bis-(2-Methoxy-4-Nitro-5-Sulphophenyl)-2H-Tetrazolium-5-Carboxanilide) (Sigma Aldrich, Germany) and PMS (Phenazine Methosulphate) (Sigma Aldrich, Germany). Actively respiring cells converted the water-soluble XTT into a water-soluble, orange-colored formazan product. The absorbance was measured at 475 nm after incubation with a microplate spectrophotometer (Benchmark Plus, BioRad, U. S. A.).

Flow cytometry and fluorescence microscopy

Flow cytometric analyses of the transfected HEK293, CHO, and A549 cells was performed to detect eGFP and CD39 expression. Therefore, 24 h, 72 h, 120 h, and 168 h after transfection, CD39-fluorescein isothiocyanate (FITC) antibody

(Abcam, U. K.: Clone A1) was used to detect CD39 expression using a FACS Calibur (BD Bioscience, U. S. A.). A total of 10,000 events were acquired in each sample. For the analysis of eGFP expression, no antibody was necessary.

The eGFP expression of CHO cells was additionally detected via fluorescence microscopy 24 h after transfection using a fluorescence microscope (IX2UCB, Olympus, Japan). The fluorescence intensity was analyzed by measuring the grey scale of the images using ImageJ.

Hemocompatibility assay

Blood-sampling procedures were approved by the Ethics Committee of the Alfred Hospital, Melbourne, Australia. Blood was collected from healthy volunteers who provided informed consent in accordance with the Declaration of Helsinki. 1 ml citrated blood was incubated with 2.5 μ l NLps, 0.5 μ g eGFP mRNA, nanoplexes (2.5 μ l NLps and 0.5 μ g mRNA), or PBS as control at 37 °C at 100 rpm. Complete blood counts were performed at different time points (2 h and 4 h) using a Hematology Analyzer (Sysmex xs-1000i, Japan).

Statistics

Unless otherwise specified, the data is represented as means with standard derivations (SD). All analysis for the two groups were performed using Student t-tests. All analyses containing more than two groups were analyzed with one-way ANOVA, comparing all groups with one another, corrected by *post hoc* Bonferroni analysis, and the corrected p-values are given. The statistical analyses were performed with the statistical software package GraphPad Prism (version 6.01, GraphPad Software, U. S. A.). Statistical significance was defined as $p < 0.05$.

Results

Generation of eGFP mRNA and CD39 mRNA

The eGFP and CD39 mRNA, and their purity, were analyzed on a 1% TBE-agarose gel (Figure 1A). The purified CD39 mRNA (1533 bp) was visualized between the 1.5 kb and 2 kb markers, and the GFP mRNA (793 bp) was visualized just below the 1 kb marker.

Characterization of cationic DC-Chol/DOPE nanoliposomes

The generated NLps had a size distribution of 110.1 ± 0.23 nm at day 1 (Figure 1B; mean \pm SD) and 106.8 ± 0.23 nm at day 30. We observed no change in size after 30 days of storage at 4 °C (Figure 1C; NS).

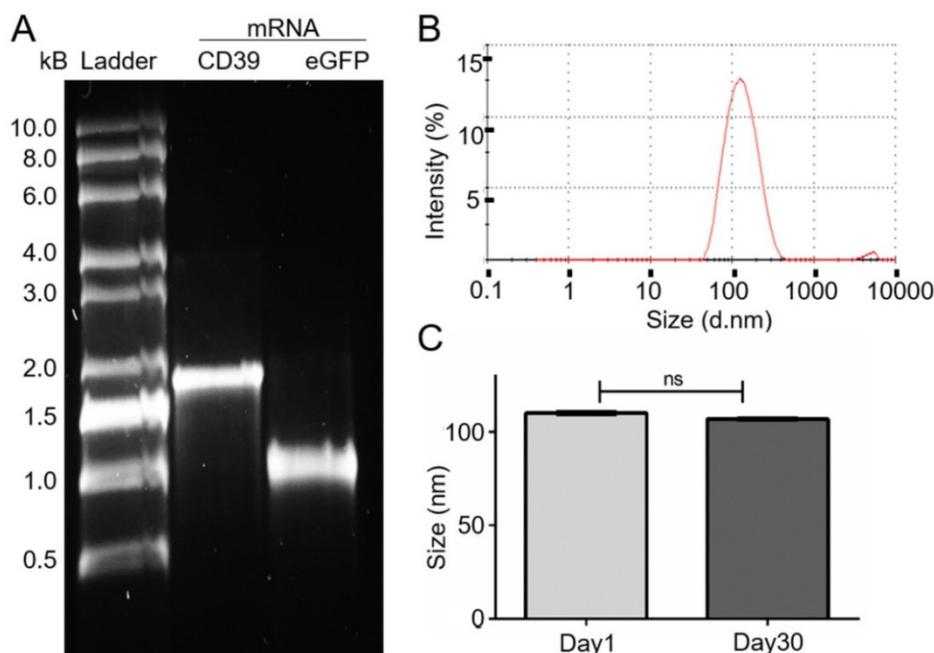


Figure 1. mRNA generation and size measurement of the DC-Cholesterol/DOPE nanoliposomes. (A) Electrophoresis with 1% agarose-TBE gel: CD39 mRNA (1533 bp), eGFP mRNA (993 bp) after purification and 0.5–10 kB RNA ladder. **(B)** Size distribution curve of the DC-Cholesterol/DOPE NLps 1 day after generation measured with a Zetazizer spectrometer exhibit an average of about 110 nm. **(C)** Comparison of size distribution of liposomes 1 day and 30 days after generation showing no difference. The two groups were compared using Student t-tests (mean \pm SD, n=5).

Evaluation of DC-Chol/DOPE nanoliposomes and nanoplexes for cell viability using CHO cells

CHO cells were transfected with either 2.5 μ l of NLps, 0.5 μ g eGFP mRNA, the same concentration of nanoplexes, or PBS as control, and an XTT-viability assay was performed at 24 h, 48 h, and 72 h post transfection (Figure 2). The transfection of liposomes showed no influence on the cell viability even 72 h after transfection with NLps, mRNA, or nanoplexes, as compared to the PBS control (102.73 ± 10.5 vs 97.95 ± 18.23 vs. 97.29 ± 8.9 vs. 99.99 ± 2.3 ; % viability mean \pm SD respectively; NS). Similar results were observed at shorter time points of 24 h and 48 h (NS). Lower doses of NLps, mRNA concentrations, or nanoplexes also showed no change in cell viability (Supplemental Figure 1).

Evaluation of transfection efficiency using nanoplexes with eGFP mRNA *in vitro*

The transfection efficiency of the liposomes was investigated 24 h after transfection with eGFP nanoplexes in a fluorescence microscope (Figure 3). There were no fluorescence signals imaged on the controls (Supplemental Figure 2). There was increased fluorescence, which was confirmed with analysis of grey-scale intensity, for cells transfected with nanoplexes (1 μ l NLps, 0.25 μ g mRNA), as compared to 1 μ l NLps, eGFP mRNA, or PBS control (16.86 ± 47 vs. 2.01 ± 1.22 vs. 1.49 ± 0.54 vs. 1.58 ± 0.64 ; mean \pm SD

respectively; AU; $p < 0.001$). This increase in the grey-scale intensity was also measured when increased amounts of NLps were used in the nanoplexes (2.5 μ l NLps, 0.25 μ g mRNA), as compared to 2.5 μ l NLps as the control (14.29 ± 5.30 vs. 1.87 ± 0.46 ; mean \pm SD; AU; $p < 0.001$).

The measurement of % eGFP expressed using flow cytometry showed an efficient transfection 24 h post incubation for nanoplexes (1 μ l NLps, 0.25 μ g mRNA), as compared to 1 μ l NLps, eGFP mRNA, or PBS control (43.57 ± 20.59 vs. 3.80 ± 2.78 vs. 1.82 ± 1.34 vs. 0.49 ± 0.30 ; % mean \pm SD respectively; Figure 4A; $p < 0.001$). This increase in the % eGFP expression was also measured when increased amounts of NLps were used in the nanoplexes (2.5 μ l NLps, 0.25 μ g mRNA), as compared to 2.5 μ l NLps as control (57.29 ± 14.70 vs. 10.56 ± 5.48 ; % mean \pm SD; Figure 4B; $p < 0.001$). The eGFP fluorescence measurement showed increased intensity with nanoplexes (1 μ l NLps, 0.25 μ g mRNA), as compared to 1 μ l NLps, eGFP mRNA, or PBS control (188.30 ± 79.17 vs. 4.7 ± 0.45 vs. 2.30 ± 0.20 vs. 3.76 ± 0.85 ; mean \pm SD respectively; AU; $p < 0.001$). Additionally, the same increase in fluorescence intensity was observed with higher amounts of nanoplexes (2.5 μ l NLps, 0.25 μ g mRNA), as compared to 2.5 μ l NLps as control (186.50 ± 42.53 vs. 5.61 ± 1.67 ; mean \pm SD respectively; AU; $p < 0.001$). Similar results were observed when Opti-MEM™ was used as the transfection medium (Supplemental Figure 3).

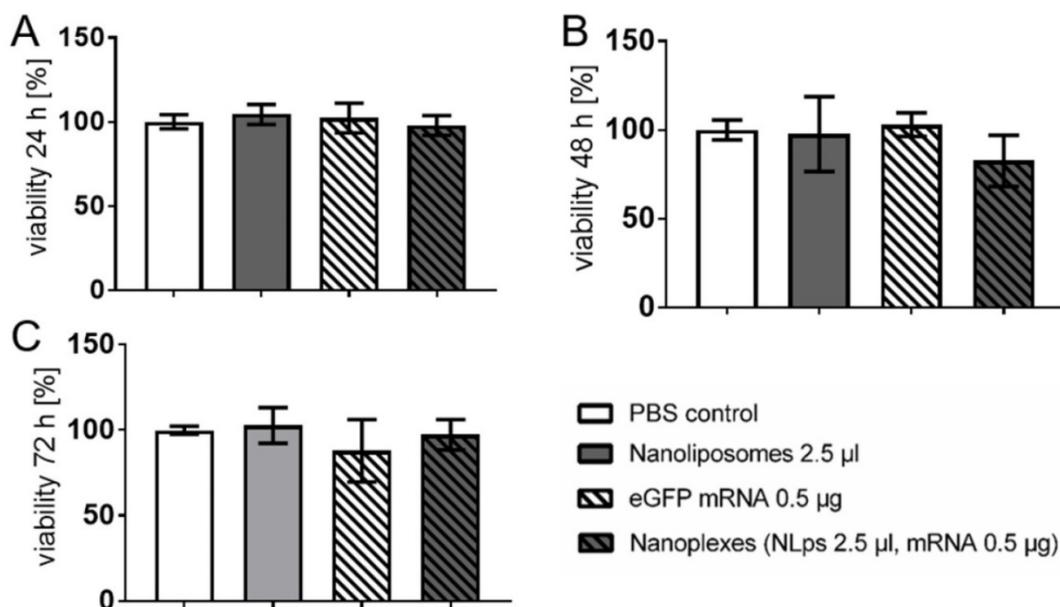


Figure 2. XTT-viability test showing no influence of the transfection of nanolipoplexes on CHO cells. CHO cells were transfected with eGFP mRNA and liposomes. The cell viability was analyzed 24 h (A), 48 h (B), and 72 h (C) after transfection using XTT-tests and OD measurement, displaying no significant difference of the nanoplexes compared to the PBS, NLps, and mRNA controls. The PBS control was set to 100% and the different groups were compared by use of repeated ANOVA measures with Bonferroni post-tests (mean \pm SD, $n \geq 3$).

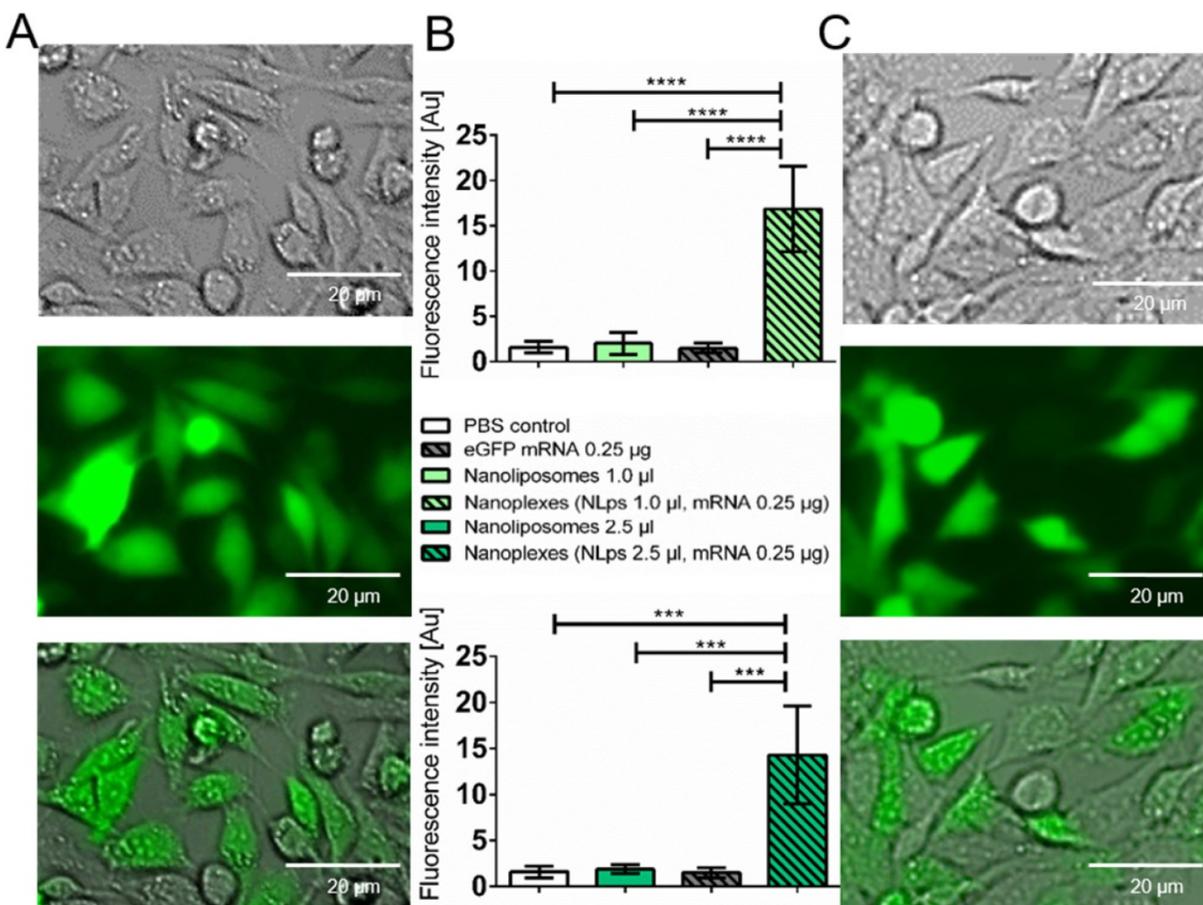


Figure 3. Microscope analysis demonstrates transfection efficiency of eGFP mRNA nanoplexes with media without FBS. CHO cells transfected with eGFP mRNA and different amounts of liposomes were analyzed 24 h after transfection under a fluorescent microscope. Representative images demonstrate a distinct expression with 1.0 µl nanolipoplexes (A) and 2.5 µl nanolipoplexes (C), supported by measurement of the fluorescence intensity via comparing the grey scale of fluorescent pictures (B). Bar charts were compared by use of repeated ANOVA measures with Bonferroni post-tests (mean \pm SD, $^{***}p > 0.001$, $^{****}p > 0.0001$).

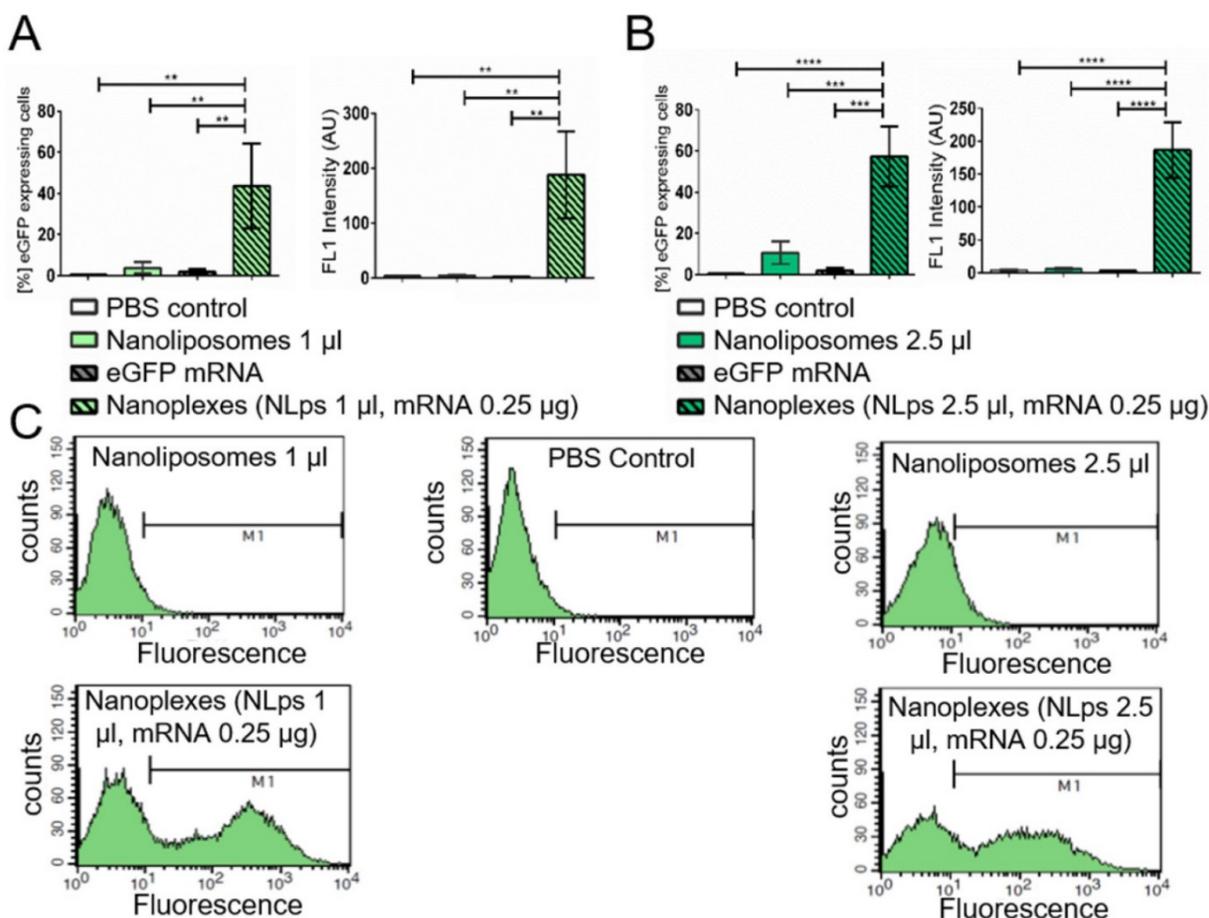


Figure 4. Flow cytometry demonstrating the transfection efficiency of eGFP mRNA nanoplexes in media without FBS in CHO cells. Significant transfection efficiency could be seen 24 h after transfection with eGFP mRNA using 1 µl (A) and 2.5 µl (B) nanoliposomes in flow cytometry. Bar graphs depict the % of protein expressing cells, as well as the median fluorescence intensity AU. The different groups were compared using repeated ANOVA measures with Bonferroni post-tests. (C) Representative fluorescence histograms are shown underneath the bar graphs (mean \pm SD, *** p <0.001, **** p >0.0001).

In vitro analysis of CD39 expression post transfection with nanoliposomes and therapeutic CD39 mRNA

The short-term transfection efficiency of the therapeutic mRNA CD39 was evaluated using CHO cells the day prior to flow cytometry analysis (Figure 5). Using an anti-CD39 FITC-bound antibody, we showed successful CD39 protein expression 24 h post incubation with nanoplexes (1 µl NLps, 0.25 µg CD39 mRNA), as compared to 1 µl NLps, CD39 mRNA, or PBS control (36.82 ± 17.74 vs. 2.91 ± 1.85 vs. 3.9 ± 0.45 vs. 1.72 ± 1.38 ; % mean \pm SD respectively; AU; p <0.01). This used higher doses of nanoplexes (2.5 µl NLps, 0.25 µg CD39 mRNA), as compared to 2.5 µl NLps as control (45.64 ± 15.31 vs. 9.11 ± 4.28 ; % mean \pm SD respectively; AU; p <0.001). Similar results were obtained using Opti-Mem™ as the transfection medium (Supplemental Figure 4).

Using an alternative cell line, we monitored for changes in cell morphology under the microscope. A higher dose of nanoplexes (5 µl NLps, 0.5 µg CD39

mRNA) was added to HEK293 cells, and we observed that there was no influence on cell morphology at both 24 h and 72 h post transfection (Figure 6A). The % of HEK cells showing CD39 protein expression 72 h after transfection with nanoplexes was significantly higher than those with NLps only or PBS control (69.5 ± 9.0 vs. 6.42 ± 1.34 vs. 0.61 ± 0.10 ; % mean \pm SD respectively; p <0.001) (Figures 6B and 6C).

When performing a longer term transfection in another cell line, the A549 cells, elevated CD39 expression was observed 120 h (5 days) and 168 h (7 days) after transfection (Figure 7). A significant increase in CD39 expression was observed at 120 h post incubation with the nanoplexes, as compared to NLps, mRNA, or PBS control (73.36 ± 6.61 vs. 8.75 ± 0.08 vs. 14.54 ± 4.41 vs. 4.42 ± 0.89 ; % mean \pm SD respectively; p <0.001). Similarly, at 168 h transfection, nanoplexes resulted in significantly increased CD39 expression as compared to controls (58.83 ± 7.65 vs. 8.27 ± 4.92 vs. 7.47 ± 2.31 vs. 5.69 ± 0.43 ; % mean \pm SD respectively; p <0.001).

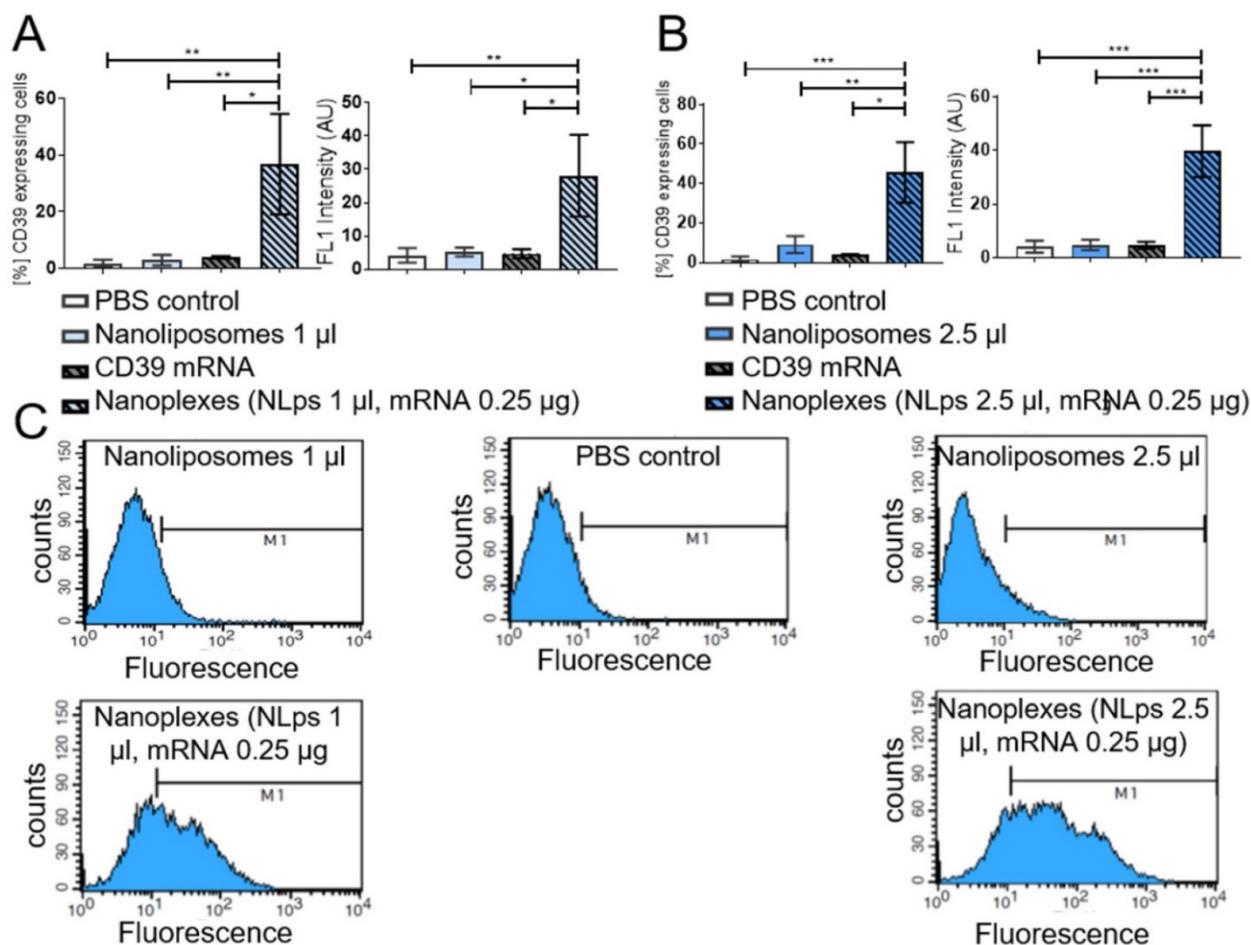


Figure 5. Flow cytometry demonstrating the transfection efficiency of CD39 mRNA nanoplexes in media without FBS in CHO cells. Significant transfection efficiency could be seen 24 h after transfection with CD39 mRNA using 1 µl (A) and 2.5 µl (B) nanoliposomes using an anti-CD39-FITC antibody in flow cytometry. The bar graphs depict the % of protein expressing cells, as well as the median fluorescence intensity AU. The different groups were compared using repeated ANOVA measures with Bonferroni post-tests. (C) Representative fluorescence histograms are shown underneath the bar graphs (mean ± SD, ** $p < 0.01$, *** $p > 0.001$).

Investigation of hemocompatibility using nanoliposomes and nanoplexes

Hemocompatibility was investigated using a hematology analyzer at 2 h and 4 h post incubation. Analyses of various cell types demonstrated no significant difference 2 h or 4 h post incubation with nanoplexes, NLps only, mRNA only, or PBS control (NS). Measurements were performed on white blood cells, red blood cells, hematocrit, platelets, lymphocytes, and monocytes (Figure 8; Supplemental Figure 5 and Supplemental Figure 6).

Discussion

We have demonstrated that NLps are suitable for the effective transfection of mRNA *in vitro*. In addition to the fact that they do not affect the viability of the cells, we have also demonstrated that they are hemocompatible. The nanoplexes, NLps, and

loaded mRNA did not alter platelet function, and hence would not result in an increased risk of thrombus formation. Using three different cell lines as target cells, we observed protein expression of CD39 for up to 7 days post transfection even with small amounts of our nanoplexes. Moving forward, this proof of concept study will allow us to develop our NLps as a nanotheranostic technology to be used in the delivery of gene therapy *in vivo*.

The concept of the delivery of mRNA to be transcribed *in vivo* as a potential new drug class to deliver genetic information has attracted much interest as an alternative to plasmid DNA transfection.^{1,18,19} Since mRNA transfection does not need entry to the nucleus, it does not integrate into the genome, thereby removing its risk of irreversible genomic modification and its transfection remains transient.

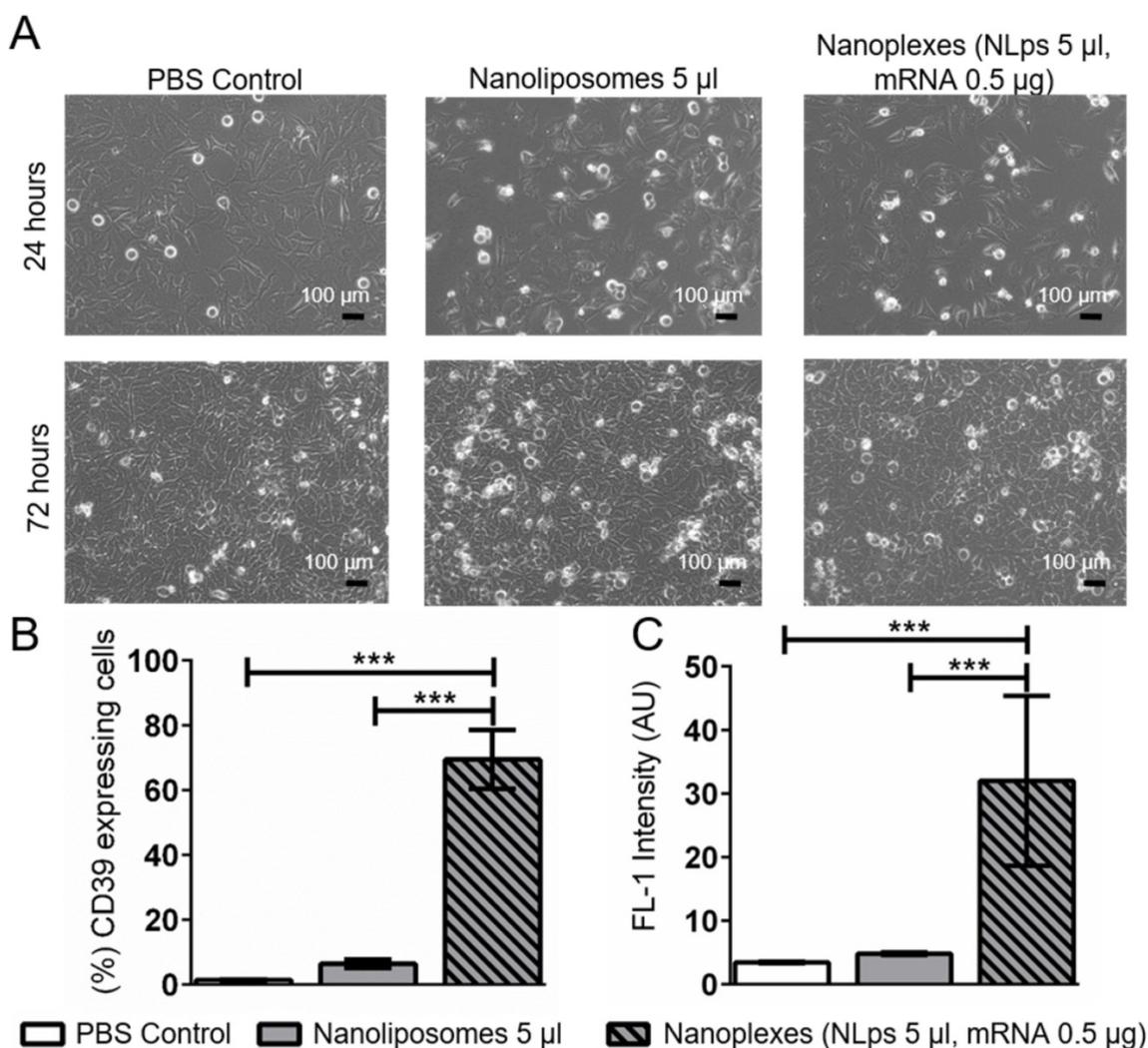


Figure 6. CD39 mRNA nanoplexes demonstrating no influence on cell morphology, growth, or transfection efficiency in HEK293 cells 72 h after transfection in flow cytometry. (A) Representative microscope pictures show no influence on morphology or growth of 5 µl of CD39 mRNA nanoplexes after transfection on HEK293 cells after 24 h and 72 h. The transfection efficiency (%) (B) and mean fluorescence intensity (AU) (C) were evaluated additionally in flow cytometry 72 h after transfection using an anti-CD39-FITC antibody. The flow cytometry assay was analyzed using repeated ANOVA measures with Bonferroni post-tests (mean ± SD, ***p>0.001).

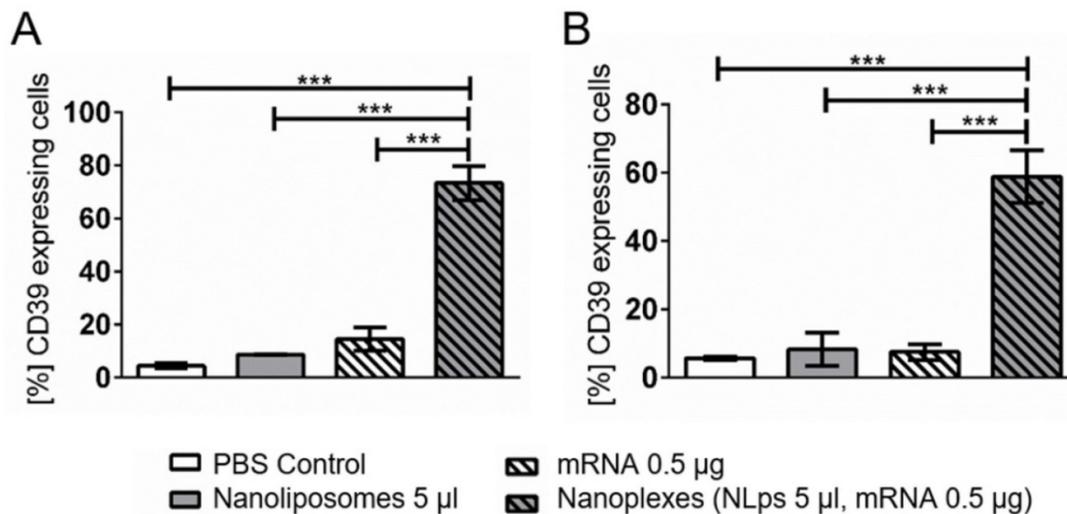


Figure 7. CD39 mRNA nanoplexes show long-term transfection in A549 cells in flow cytometry. CD39 mRNA and 5 µl of nanoplexes were transfected in A549 cells. The efficiency was analyzed in flow cytometry 120 h (A) and 168 h (B) after transfection using an anti-CD39-FITC antibody. The bar graphs depict the % of protein expressing cells. These assays were analyzed using repeated ANOVA measures with Bonferroni post-tests (mean ± SD, ***p>0.001).

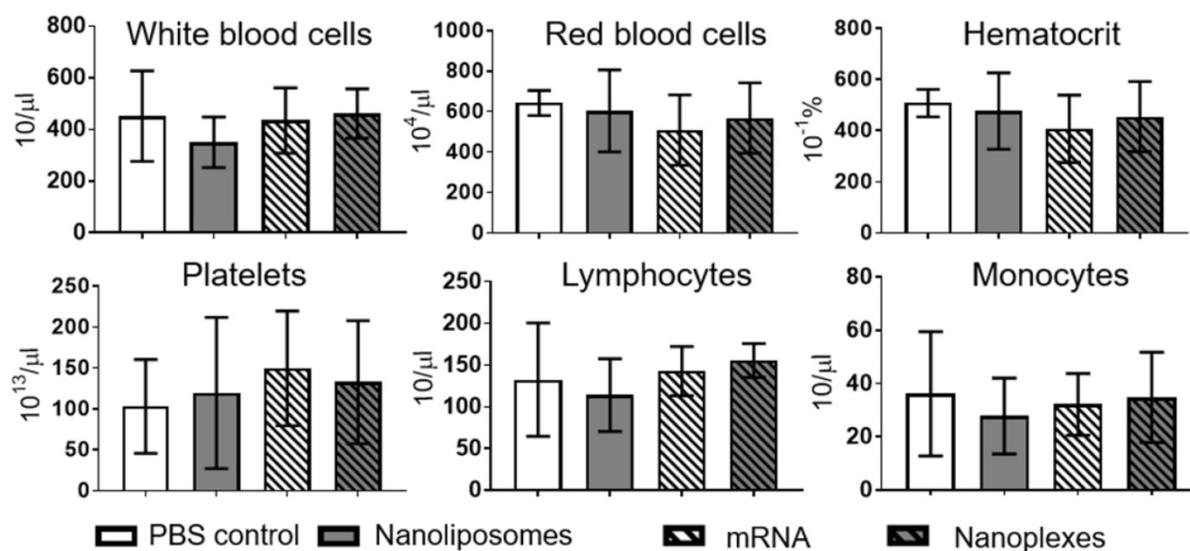


Figure 8: Hemocompatibility analysis demonstrates no effects of nanoliposomes or nanoplexes on human whole blood after 2 h. The complete blood count was analyzed after 2 h of incubation of whole human blood with nanoliposomes, mRNA, and nanoplexes at 37 °C for 4 h in total. No significant difference was seen in white blood cells, red blood cells, hematocrit, platelets, lymphocytes, or monocytes. The bar graphs were analyzed using repeated ANOVA measures with Bonferroni post-tests (mean ± SD, n=5).

Many studies have already shown that, in comparison with DNA transfection, mRNA can be transfected and translated efficiently, and that mRNA-based therapy exhibits several advantages.^{16,20,21} The cytoplasmic expression system utilized by mRNA enables quiescent or slower proliferating cells, including hepatocytes, vascular endothelial, and muscle cells, to be transfected.¹⁹ However, Barreau et al. observed that large amounts of free RNA molecules could become toxic and trigger stress responses of cells.²² Another concern is that the half-life of mRNA might be limited, as the mRNA might degrade after being added *in vitro* or *in vivo*, and so the ability for transfection and desired protein production might only last a few days.²² An elegant study by Rejman et al. demonstrated mRNA transfection of up to 9 days using 2 μg of mRNA.²³ In our study, we have observed expression of CD39 for 7 days post incubation with our nanoplexes containing only 0.5 μg of mRNA; therefore we have not observed any toxicity. As compared to other studies like that of Zohra et al., who used 3 μg mRNA, we have shown a high transfection efficiency using as little as 0.25 μg mRNA.²⁴

Several methods have been used for the transfection of mRNA, mainly classified into viral and non-viral delivery.^{1,18,19} Viral delivery is more costly because the generation of such vectors for clinical use will likely be expensive. Electroporation as a method of transfection has been studied intensively, and used mainly for *in vitro* work. There have also been concerns in regard to the damage that electroporation may cause to the cells. Therefore, the ideal gene

delivery carrier should guarantee a secure gene transport without exhibiting any immune response or toxicity, and should be able to be synthesized in a large scale.²⁵ In order to diverge from the use of viral vectors, chemical transfections using non-viral carriers, such as cationic liposomes, polycationic polymers, nanoparticles and cationic dendrimers have been extensively studied.^{25,26} Of these, the use of cationic liposomes is the best established vehicle and it provides a cost-effective and safe method.⁶ The commercially available lipofectamine is widely used for DNA, mRNA, and siRNA transfection; however, it comes with cell toxicity.²⁷ The protocol for incubation with lipofectamine is therefore usually less than 4 h.⁴

Our NLps and nanoplexes did not lyse the cells in blood, nor cause any thrombotic events. In addition, our *in vitro* assays showed good cell viability even after long-term incubations with NLps or nanoplexes. In addition, many studies have observed that Opti-MEM™ must be used for positive nucleic acid transfection because of interference with the nanocomplex formation.^{12,20} In our study, we have used Opti-MEM™ as our control, but have also successfully demonstrated equal efficiency in transfection using the cell lines' preferred media without FBS.

Our cationic NLps consist of the cationic lipid DC-Cholesterol and the neutral helper lipid DOPE. This combination has been presented in several transfection studies.^{15,28,29} The positively charged head group of DC-Cholesterol results in the delivery of genetic materials into various cell lines; the incorporation of cholesterol into liposomes has also

been shown to prevent aggregation, improve serum stability, and enhance transfection efficiency.^{10,14,17} Neutral lipids like DOPE increase the rigidity and delivery efficiency, and to aid in the assembly of cationic liposomes.^{11,15,30}

If the circulation half-life or the size of the NLps need to be prolonged, they can be stabilized using polyethylene glycol (PEG). The incorporation of PEG corona might protect them against degradation, and this modification may in turn enable a longer transfection and greater protein expression to be achieved. In addition to their transfection ability, there is great potential for the development of these NLps for co-delivery of other therapeutic drugs or imaging agents.¹⁹ In the field of cancer, where Zong et al. demonstrated stable and efficient delivery via pH-sensitive polydopamine-coated liposomes loaded with the chemotherapeutic agent 5-fluorouracil (5-FU).³¹ Wang et al. presented a cationic PLGA-folate-coated PEG-liposome for targeted co-delivery of the drug doxorubicin, along with DNA gene therapy, in MDA-MB-231 tumor cells.³²

The development of effective nanocarriers and the improvement of their delivery system play important roles in the detection of biomarkers and treatment of diseases. Within the past years, many have recognized that liposomal delivery of drugs and nucleic acids is one of the most interesting theranostic approaches for safe gene transfection and therapy.^{7,15,33} Efforts have been invested to increase the stability of liposomes, as well as superior encapsulation, greater biocompatibility, and higher loading efficiency for better biodistribution, enabling them to be employed as theranostic agents in clinical studies.^{24,29,34,35}

The clinical translation of the described therapeutic approach is feasible. Several published studies have shown the clinical applicability of mRNA vaccination.³⁶⁻³⁹ Pollard et al. reported that mRNA encoding of the HIV gag antigen, complexed with DOTAP/DOPE liposomes, could be successfully delivered to DC cells for immunization.³⁷ This approach also showed reduced innate immunity, compared to initial performances of DNA vaccines. In another promising study based on Novartis's proprietary adjuvant MF59 combined with the lipid DOTAP, a cationic emulsion was employed for the stimulation of the immune response via the antigen-specific CD4⁺ T-cell response.⁴⁰

Perspective for the transfection of CD39 mRNA

In our study, we have demonstrated the successful protein expression of CD39 in three different cell lines after mRNA transfection. This

clearly demonstrates the potential for an anti-thrombotic and anti-inflammatory genetic therapy. CD39 is an ectonucleoside triphosphate diphosphohydrolase which degrades adenosine 5'-diphosphate (ADP), a major player in the platelet activation cascade, and therefore represents a promising anti-thrombotic strategy.⁴¹ In the vasculature, CD39 is normally expressed on the surface of endothelial cells, preventing platelets from activation and attachment to resting endothelium. Therefore, CD39 plays an important role in response to vascular injury and homeostasis, and plays an essential role in modulating vascular thrombosis and inflammation.⁴² In fact, several studies have shown the benefits of elevated CD39 levels in animal models of thrombosis, including myocardial infarction and stroke.^{42,43} In addition, based on the fact that NTPDase activity is substantively lost in the vascular system of injured or rejected grafts, Robson et al. described the potential of targeted expression of CD39 in transplantation-associated application and thrombosis.⁴⁴ However, similar to current anti-thrombotic drugs, this therapy is associated with concentration-dependent bleeding complications.^{45,46} To overcome this, Hohmann et al. also created an activated platelet-specific single-chain antibody fused with the CD39 protein and demonstrated the anti-thrombotic effects of this fusion construct in a carotid artery thrombosis model.⁴⁷

Likewise, the use of our NLps for transfection of CD39 mRNA will allow the transmembrane protein to be expressed on the cell surface. The hydrolyzing effects of CD39 expressed on the cell will in turn prevent inflammation and also provide anti-platelet activity, both of which will hinder the development of atherosclerosis and thrombosis.^{43,44} The current proof of concept study will be the basis for further modification of our NLps for targeted therapeutic gene delivery and possible cell specific transfection to prevent platelet activation and inflammation. However, as is the case for nanotechnology in general, the clinical utility of our approach remains to be proven.

Conclusion

Recent developments in the field of liposomal delivery of therapeutic nucleic acids provide a wide range of applications and improved therapeutics. We have generated NLps using cationic DC-Cholesterol and DOPE for the delivery and transfection of therapeutic mRNA into three cell lines. We have observed no effects on cell viability or hemocompatibility. The finding that CD39 protein expression was evident for 7 days indicates that this non-viral gene delivery carrier could potentially be

used for long term transfections in various areas of medical need. Such a nanotheranostic approach would be of particular interest for areas such as anti-inflammation, anti-thrombotic and anti-cancer therapy.

Abbreviations

CD39: ectonucleoside triphosphate diphosphohydrolase-1; CHO: Chinese hamster ovary; DC-Chol: 3 β -[N-(N',N'-dimethylaminoethane)-carbonyl]cholesterol; DMEM: (Dulbecco's Modified Eagle's medium); DOPE: 1,2-dioleoyl-sn-glycero-3-phosphoethanolamine; DOTMA: 1,2-di-O-octadecenyl-3-trimethylammonium propane; eGFP: enhanced green fluorescence protein; FACS: fluorescence activated cell sorting; FBS: fetal bovine serum; HEK293: human embryonic kidney cells; mRNA: messenger ribonucleic acid; NLps: nanoliposomes; OD: optical density; PMS: phenazine methosulphate; siRNA: small interfering RNA; XTT: 2,3-Bis-(2-Methoxy-4-Nitro-5-Sulfophenyl)-2H-Tetrazolium-5-Carboxanilide.

Supplementary Material

Supplementary figures.

<http://www.ntno.org/v01p0154s1.pdf>

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Competing Interests

The authors have declared that no competing interest exists.

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